

Best practice skin antisepsis for insertion of peripheral catheters

Simon Clare and Stephen Rowley

Recent healthcare modelling based on published data has estimated 653 000 healthcare-associated infections (HAIs) annually among adult inpatients in English NHS hospitals, with 22 800 annual deaths (Guest et al, 2020). Collectively, the cost to the NHS in England of these challenging numbers is a staggering £2.7 billion. Bloodstream infections represent an estimated 7.3% of the total HAIs (Guest et al, 2020) with as many as 70% of catheter-associated bloodstream infections (CABSIs) thought to be preventable (Umscheid et al, 2011). Peripheral intravenous catheters (PIVCs) are the most frequently used invasive devices in hospitals with estimates as high as 70% of all inpatients requiring a PIVC during their stay in the hospital environment (Zingg et al, 2009). Based on NHS Supply Chain data, more than 300 English NHS trusts purchase more than 25 million safety peripheral intravenous catheters (SPIVCs) annually (NHS Clinical Evaluation Team, 2018).

Growing concern for PIVC care

Notwithstanding concern regarding unnecessary insertion, recent research has highlighted the under-reported risks posed by the placement and management of PIVCs including inadequate technique for skin antisepsis (Zhang et al, 2016; Mermel, 2017; Saliba et al 2018; Blanco-Mavillard et al, 2019; Høvik, 2019).

Although superficially PIVC infections look to be smaller in number than central venous access device (CVAD) infections, considerably more PIVCs are inserted overall compared with CVADs and absolute infection rates are in fact similar for both device types (Zhang et al, 2016; Sato et al, 2017). Moreover, there is a high rate of reported PIVC failure (Helm et al, 2015). In the state of Pennsylvania in the USA, most untypically, hospitals are required to report all laboratory-confirmed bloodstream infections (LCBIs), not solely the more typical central-line associated bloodstream infections (CLABSIs). Data from 2011–2012 highlighted the large, and increasing, numbers of line bacteraemia that were not CLABSIs, raising the question: how many were actually PIVC infections (Davis, 2014)? There is nothing to suggest that this might not be a typical incidence, when reported, and raises a question of effective surveillance. All these authors, and others, highlighted that to improve patient safety, there is a compelling need to focus anew on PIVC insertion and maintenance (Trinh et al, 2011).

As the risks of PIVC infection are better recognised there has been a transition towards longer PIVC dwell times. For some

ABSTRACT

This article discusses the importance of effective skin antisepsis prior to the insertion of peripheral intravenous catheters (PIVCs) and how best clinical practice is promoted by application of an appropriate method of skin disinfection integrated effectively with a proprietary aseptic non touch technique, or other standard aseptic technique. Historically under-reported, incidence of infection and risk to patients from PIVCs is now increasingly being recognised, with new research and evidence raising concern and helping to drive new clinical guidance and improvement. The risks posed by PIVCs are particularly significant given increasing PIVC dwell times, due to cannula removal now being determined by new guidance for clinical indication, rather than predefined time frames. Clinical 'best practice' is considered in context of the evidence base, importantly including availability and access to appropriate skin antisepsis products. In the UK, and other countries, ChloraPrep is the only skin antisepsis applicator licensed as a drug to disinfect skin and help prevent infections before invasive medical procedures, such as injections, blood sampling, insertion of PIVCs and minor or major surgery.

Key words: ANTT ■ Aseptic technique ■ Skin antisepsis ■ ChloraPrep

years, dwell times have remained within predefined time frames, varying between 24 hours and 96 hours. However, the advent of 'clinical indication', based on best evidence, recommends the removal of PIVCs based on site and device integrity—notably, this is open-ended (Van Donk et al, 2009; Rickard et al, 2010; Webster et al, 2019). This presents both cause for potential concern and opportunity for improvement. In other words, although this evidence-based approach to PIVC removal is welcomed, it also highlights the need for improved standards of PIVC insertion, maintenance and surveillance, and offers scope for innovation.

Skin and skin flora in context of PIVCs

Human adult skin is approximately two square metres in area and weights around 3.6 kg. This important organ is a primary defence against pathogenic microorganisms. Its structure and purposes are well understood; however, perhaps less understood

Simon Clare, Research and Practice Development Director ANTT, The Association for Safe Aseptic Practice, simon.clare1@nhs.net

Stephen Rowley, Clinical Director ANTT, The Association for Safe Aseptic Practice, stephen.rowley@annt.org

Accepted for publication: November 2020

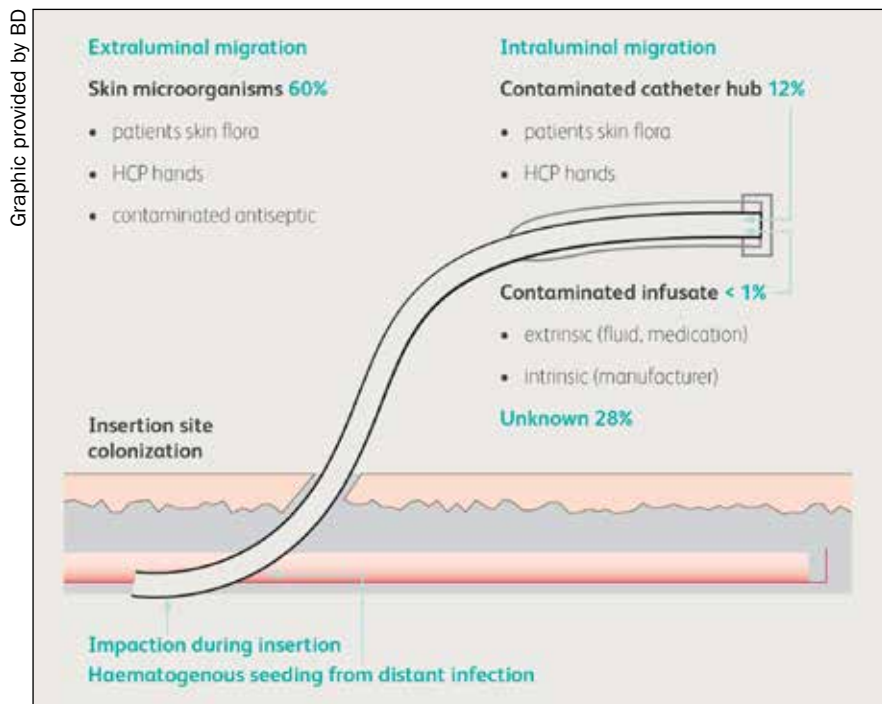


Figure 1. Routes of infection in peripheral intravenous catheters (adapted from Safdar and Maki, 2004)

is the microbiology of skin flora and the potential for infection when breached or disturbed by PIVC access. An area of around 1 cm² of skin can host as many as 10 million aerobic bacteria, which are a leading cause of HAIs (Hibbard, 2005). Importantly, because approximately 80% of skin flora resides in the first five cell layers of skin (Brown, 1989), when the skin is incised or cannulated, the exposed tissue is at risk of contamination, which can lead to extraluminal microorganism migration through the created ‘wound’, potentially causing infection, and secondary biofilm development caused by intraluminal migration later in the process (Zhang et al, 2016). Each and every time a vascular access device (VAD) is inserted, accessed or managed (applying or changing a PIVC dressing), failures in aseptic technique leave the patient vulnerable to microbial migration from both their own skin and the healthcare environment (the health professional, the physical environment and the air environment).

In this light, the skin is both an important protector and protagonist in prevention of healthcare-associated infections (HAIs). In the USA, it has been estimated that 60% of CABSIs are caused by micro-organisms from the patient’s skin (Safdar and Maki, 2004; Maki et al, 2006) (Figure 1). Numerous microorganisms are considered a part of the normal human skin flora, including *Staphylococcus epidermidis* and *Staphylococcus aureus*. It is worth noting that PIVCs are one of the most common sources of *Staphylococcus aureus* bacteraemia (Blauw et al, 2019).

Best practice in skin antiseptics

Effective skin antiseptics (see Box 1) plays a critical role in protecting patients from infections during invasive clinical procedures, particularly during the placement of indwelling VADs (Zhang et al, 2016). To mitigate risk, best practice can

be defined as ‘quality care deemed optimal based upon a prevailing standard described by evidence’ (Nelson, 2014). For skin antiseptics, best practice includes an appropriate application technique (Casey et al, 2017), a time period for application and complete drying (Tepus et al, 2008; Silva, 2014), plus actions and methodologies to minimise the risk of introducing harmful microorganisms (Rowley et al, 2010) into the procedure.

Best practice naturally includes an appraisal of the most effective disinfection solution and method of application. In recent years, based on best evidence, clinical guidelines have increasingly advised the use of chlorhexidine gluconate in combination with isopropyl alcohol as being most effective in reducing CABSIs (Loveday et al, 2014; Mimoz et al, 2015; Gorski et al, 2016).

Probably reflecting the robust constitution of the skin and its various layers, it would seem that the manner of application of skin disinfectant is significant too. A ‘cross-hatching’, back-and-forth disinfecting technique is considered to be 10 times more effective at reducing bacterial load than the traditional so-called concentric circle technique for skin antiseptics (McDonald et al, 2001). It enables maximum contact between the skin and antiseptic, and helps the solution to reach and disinfect deeper cell layers of the skin (Silva, 2014). Traditionally, the concentric circle technique has persisted in practice without scientific evidence to warrant its use (Tepus et al, 2008; Hadaway, 2012; Tung, 2013). Therefore, recommended skin antiseptics technique for PIVC insertion is a combination of a ‘cross hatch’, disinfection technique, with 2% chlorhexidine gluconate in 70% isopropyl alcohol, then allowing the site to air dry prior to insertion. For patients with sensitivity to chlorhexidine, povidone iodine in alcohol can be used (Tepus et al, 2008; Hadaway, 2012; Tung, 2013; Loveday et al, 2014; Silva, 2014; Gorski et al, 2016; Casey et al, 2017).

The Association for Safe Aseptic Practice (ASAP) works to support improvement in standards of aseptic technique using the ANTT Aseptic Non Touch Technique, a proprietary comprehensive clinical practice framework for aseptic technique. Fit-for-purpose medical equipment and supplies are naturally a key factor in achieving effective ANTT, or indeed any type of aseptic technique. To this end, the ASAP’s work includes supporting industry to help ensure synergy and safety between products and aseptic practice. Medical products can, in fact, bring about significant improvement in aseptic technique when novel designs address problematic human factors. A good example of this has been the advent of passive disinfection for IV hub disinfection that reduced reliance on specific cleaning techniques (Moureau and Flynn, 2015; Gorski et al, 2016). A similar step change has been seen for skin antiseptics with the advent of purpose-designed applicators.

Product types for PIVC skin antiseptics

The two most common product types used to deliver chlorhexidine/isopropyl alcohol skin disinfection for skin PIVC are fibre-based ‘wipes’ and specifically designed for purpose hand-held applicators. Currently, the only single-use applicators that are licensed and commercially available are BD ChlorPrep™ (Beckton, Dickinson, UK) applicators. ChlorPrep is licensed by the Medicines and Healthcare products Regulatory Agency

(MHRA), the UK's licensing regulator that evaluates safety and efficacy of medicines and medical devices. ChloroPrep is specifically licensed as a medicinal product intended for preoperative skin antisepsis. There is an assumption that health professionals will adhere to MHRA guidance by using the appropriate licensed medicinal product for preoperative skin antisepsis. MHRA guidance does indeed state that using the appropriately authorised product for its specific intended use is the best way of minimising harm. In practice, MHRA guidance is not to be taken as a complete or definitive statement of the law. As such, there is nothing to restrict a health professional from using non-licensed chlorhexidine products for skin preparation prior to invasive procedures such as the insertion of PIVCs. Of note, there is guidance provided by professional bodies such as the General Medical Council (GMC) and Nursing and Midwifery Council (NMC) on the use of an unlicensed product as a medicine. Despite this, some products that the MHRA would classify as medical devices or even biocides could be, and in some cases are, used for skin antisepsis.

This situation does not necessarily present patient risk for preoperative skin antisepsis, but quite simply, the risks compared with licensed products are unknown. The MHRA, as with similar regulatory bodies in other countries, ensures that products prove their efficacy and safety before patients are exposed to them. When unlicensed products are used for skin antisepsis this regulatory structure is bypassed and healthcare organisations, and not least patients, have no assurance that the product they are using has undergone stringent testing, and will also not have a formal mechanism for surveillance and feedback of any adverse incidents.

Organisations that accept this risk for financial or other rationale typically acknowledge their use of unlicensed products as a medicine on a local risk register. In light of the Mid-Staffordshire Public Inquiry and the call for greater transparency in healthcare organisations (Francis, 2013), one might consider how transparent in actual practice, this approach is to patients. Either way, the MHRA is unequivocal: 'Where an authorised product exists this should be used in preference to another product...' (MHRA, 2020).

ChloroPrep

ChloroPrep applicators address a number of problematic issues and human factors for skin antisepsis. They contain a sterilised product solution of chlorhexidine gluconate and isopropyl alcohol, addressing the potential negative impact of using non-sterile solution for skin antisepsis (Vigeant et al, 1998; Weber et al, 2007; Leong et al, 2018; Song et al, 2018; , 2019). This risk led the US medicines regulatory body, the Food and Drug Administration (FDA), to make it mandatory for manufacturers to write on the packaging whether the content of a skin antisepsis product is sterile or not (FDA, 2016).

ChloroPrep products contain a defined and visible amount of chlorhexidine/isopropyl alcohol solution with assurance of stability ensuring sufficient fluid volume to counter evaporation and facilitate adequate skin coverage and skin impregnation (Tarka et al, 2019). Different volumes of solution are required for different procedures and ChloroPrep products range from 0.67 ml

for peripheral application to 26 ml for surgical procedures. In contrast, wipes typically do not appear to assure a precise volume per wipe, seemingly creating uncertainty in usage.

Wipes can be used with effective non-touch technique; however, the various possible approaches to their use increase potential for human factors and subsequently practice variability. For example, wipes may be used folded, unfolded or 'scrunched up'. Such variables can impact effective non-touch technique. Wipes also bring the operator's fingers closer to the Key-Site. This is not addressed in product instruction because wipe manufacturers, of course, do not provide detailed instructions for their use for skin antisepsis as this is not their intended or indeed stated function. (They are typically stated for disinfecting medical devices or helping remove adhesive residue from dressings.) Method of use can therefore be unclear and left open to interpretation—for example, how and how much the wipe should be unfolded and how best handled to ensure a non-touch technique.

ChloroPrep applicators are opened ready for use, albeit requiring an action to first release the solution. Hand-held wands then promote non-touch technique by separating the health professional's fingers from the applicator surface and the insertion site, helping to protect Key-Parts and Key-Sites (see *Box 1*). A wand applicator system allows a better control of the flow rate being delivered onto the skin. There is essentially only one way of holding the applicator, promoting ease of use and less potential for variability. Because intended for PIVC insertion and other procedures, an applicator system is supported by manufacturer's instructions.

Until a wipe is granted marketing authorisation from the MHRA, ChloroPrep as a medicinal product is the only licensed and commercially available applicator for disinfection of the skin prior to skin antisepsis prior to invasive medical procedures in

Box 1. Glossary of terms

Aseptic technique

A set of infection prevention actions aimed at protecting patients from infection during invasive clinical procedures and management of indwelling medical devices. Asepsis is the absence of pathogenic organisms in sufficient quantity to cause infection and is achievable through aseptic technique (Gorski et al, 2021).

A N T T Aseptic Non Touch Technique

A specific and comprehensively defined type of aseptic technique with a unique theory-practice framework based on an original concept of Key-Part and Key-Site protection; achieved by integrating standard precautions such as hand hygiene and personal protective equipment (PPE) with appropriate aseptic field management, non-touch technique and sterilised supplies. It is designed for all invasive clinical procedures and management of invasive medical devices (Gorski et al, 2021).

Skin antisepsis

Antisepsis relates to techniques and practices for the removal, or elimination, of transient microorganisms from the skin and a reduction in the resident flora (Dockery, 2012).

Wipe

An impregnated fibre-like material used to disinfect skin and surfaces with the goal of removing harmful microorganisms.

Applicator

An intentional design incorporating a method of delivering skin disinfectant locally without directly touching the impregnated sponge or surface of the device.

KEY POINTS

- Skin antisepsis is a critical part of peripheral intravenous catheter (PIVC) insertion and the integration of aseptic technique is fundamental to effectiveness. Skin antisepsis applicators support more effective aseptic technique by design. The proprietary ANTT approach supports more explicit education and guidance
- Incidence of PIVC complications, including serious infection, has historically been under-reported. Improved surveillance and competency-based education and training will improve the delivery and monitoring of safe and effective practice. The successes of clinical care bundles for central venous catheters should be applied to PIVC to help mitigate problems with insertion and maintenance
- There are clear advantages for healthcare organisations in using licensed medicinal products for skin antisepsis. The use of non-licensed products potentially compromises patient safety, safety surveillance is seriously curtailed and the integration of effective aseptic technique is ambiguous

the UK. Sepp™, Frepp™ etc received an MHRA marketing authorisation for medical use and drug approval from the US FDA. However, from a small convenience sample of policy and procedure documents from NHS trusts obtained online using a Google search, only 50% (5/10) carried an instruction to use a specific product that is licensed as a medicinal product (identified as either ChloroPrep or [ChloroPrep] Sepp); 30% (3/10) did not mention the type of product, whereas 20% (2/10) specified the use of a (non-licensed) ‘wipe’ for skin antisepsis.

Case study 1. Addressing problems of practice with a bundle approach

Identifying problems with increasing incidence of PIVC infection, a bundle approach was implemented at Methodist Hospitals (Gary, Indiana, USA) based on evidence-based standards of practice (Gorski et al, 2016). The bundle integrated several practice recommendations including the use of ChloroPrep for skin antisepsis. Recognising the lack of surveillance around PIVC insertion and management, the bundle included careful assessment of the insertion site, ongoing surveillance of process and outcomes, and a review in ‘real time’ of any infections on the ward. After 12 months DeVries et al (2016) reported a 37% reduction in primary bacteraemia, a 19% reduction in PIVC-associated bloodstream infections after 24 months and a 75% reduction in CLABSI in the intensive therapy unit; this was coupled with improved patient satisfaction, longer indwell times and significant cost savings from rationalising supplies (DeVries et al, 2016).

Case study 2. The risk of suboptimal skin antisepsis

Registered nurses undergoing PIVC training in a large teaching hospital in England were asked to use and discuss the application of wipes and ChloroPrep. There was group uncertainty whether the wipe was best unfolded, left folded, or ‘scrunched up’. Some users found it difficult to only handle one side of it, with the other side exclusively in contact with the patient’s skin (non-touch technique). Towards the end of the 30-second

skin cleaning some users felt the wipe was running dry, with the skin site accordingly drying. When using ChloroPrep Sepp, some users commented on the relatively large amount of solution available and easily applied to skin. There was no debate or variance as to the handling of the device, probably due to the simple applicator design. Non-touch technique was achieved naturally by design without the need for any noticeable consideration of handling technique.

Improving practice and outcomes

Procedure-based ‘care bundles’ can help ensure that busy staff have easy access to the most appropriate equipment for any given procedure and, in effect, help direct best practice. When used for PIVC insertion and maintenance they have been shown to reduce PIVC-related bloodstream infections (Mestre et al, 2013; DeVries et al, 2016). ChloroPrep has been reported in several studies of clinical care bundles; notably, Steere and colleagues reporting results from the PIV5-Rights bundle, demonstrated improved single catheter dwell times (89% vs 15%) and reduced failure rates due to complications (11% vs 85%) (Steere et al, 2019).

To ensure patient safety, the various equipment contained in a care bundle or ‘cannulation pack’ requires effective integration with a standard aseptic technique. To this end, the widespread adoption of the proprietary ANTT approach as a standard aseptic technique in the NHS, and a de facto standard internationally (Rowley and Clare, 2020), provides a ready-made educational and clinical practice platform for using care bundles as part of PIVC improvement initiatives. Examples of this greater synergy include the inclusion of the proprietary ANTT as part of clinical care bundles to improve incidence of CRBSI (Mutalib et al, 2015; Taylor et al, 2017). Further enhancing this integration, the proprietary ANTT clinical procedure guidelines (picture-based, sequenced and risk-assessed step-by-step guidance) are a tangible and accessible translation of the proprietary ANTT approach into real-world practice; these simple but effective tools have proved extremely popular, allowing the explicit description of correct utilisation of medical devices with safe aseptic practice (Rowley et al, 2010; Rowley and Clare, 2019).

Conclusion

Skin antisepsis is a critical component of PIVC insertion and maintenance. Best practice requires the use of the proprietary ANTT approach, or other type of standard aseptic technique, to achieve and maintain asepsis during the insertion and maintenance of invasive medical devices. Compared with central venous access, there continues to be a lack of appreciation of the risks associated with insertion and maintenance of PIVCs. This is especially concerning, considering the sheer volume of PIVC insertions generally, and the significant potential for incidence of complications including bacteraemia, especially in light of the lack of surveillance for PIVC infection.

Not least, improved standard surveillance for PIVC infection would enable healthcare organisations to make more informed choices on product procurement for skin antisepsis that take into account the full cost of PIVC infection. Put differently, using unlicensed medical devices may not be cost-effective when set against the ever-increasing costs of treating bacteraemia,

antimicrobial resistance, extended hospitalisations and the realities of litigation in an increasingly litigious society.

It may be prudent for healthcare organisations to fully consider the implications of a risk-register approach to using non-licensed products, especially when factoring in the lack of surveillance, the cost of treating infection, increases in length of admissions, potential litigation and reputation. At the very least, there would appear to be room for more transparency regarding keeping patients informed on the use of unlicensed products as a medicine.

Ways to improve the safety of this most common invasive procedure for patients include effective staff education regarding the risk of PIVCs, access to appropriate licensed products for skin antisepsis, and training in integrating effective skin antisepsis with best-practice the proprietary ANTT approach for PIVC insertion. **BJN**

Declaration of interest: This article was supported by Becton, Dickinson and Company (BD); the authors were commissioned to write this article and will receive an honorarium. The authors work in the NHS and are also non-salaried board members of The Association for Safe Aseptic Practice (ASAP) a non-profit non-governmental organisation.

Blanco-Mavillard I, Rodríguez-Calero MÁ, de Pedro-Gómez J et al. Incidence of peripheral intravenous catheter failure among inpatients: variability between microbiological data and clinical signs and symptoms. *Antimicrob Resist Infect Control*. 2019; 8:124. <https://doi.org/10.1186/s13756-019-0581-8>

Blauw M, Foxman B, Wu J, Rey J, Kothari N, Malani AN. Risk factors and outcomes associated with hospital-onset peripheral intravenous catheter-associated *Staphylococcus aureus* bacteremia. *Open Forum Infect Dis*. 2019; 6(4):ofz111. <https://doi.org/10.1093/ofid/ofz111>

Brown E, Wenzel RP, Hendley JO. Exploration of the microbial anatomy of normal human skin by using plasmid profiles of coagulase-negative staphylococci: search for the reservoir of resident skin flora. *J Infect Dis*. 1989;160(4):644-50. <https://doi.org/10.1093/infdis/160.4.644>

Casey AL, Badia JM, Higgins A. Skin antisepsis: it's not only what you use, it's the way that you use it. *J Hosp Infect*. 2017; 96(3):221-222

Davis J. Peripheral vascular catheter-related infection: dwelling on dwell time. *Pennsylvania Patient Safety Advisory*. 2014;11(1):30-35. http://patientsafety.pa.gov/ADVISORIES/Pages/201403_30.aspx

DeVries M, Valentine MJ, Mancos P. Protected clinical indication of peripheral intravenous lines: successful implementation. *Journal of the Association for Vascular Access*. 2016; 21(2): 89-92. <https://doi.org/10.1016/j.java.2016.03.001>

Dockery GD. Chapter 7 – Aseptic techniques. In: Dockery GD, Crawford ME (eds) *Lower extremity soft tissue & cutaneous plastic surgery*. 2nd edn. W.B. Saunders. 2012

Food and Drug Administration. FDA Drug Safety Communication: FDA requests label changes and single-use packaging for some over-the-counter topical antiseptic products to decrease risk of infection. 2016. <https://www.fda.gov/drugs/drugsafety/ucm374711.htm> (accessed 25 November 2020)

Francis R. Report of the Mid Staffordshire NHS Foundation Trust Public Inquiry: Executive summary. 2013 (archived). <https://tinyurl.com/y5cykkff> (accessed 25 November 2020)

Gorski LA, Hadaway L, Hagle M, McGoldrick M, Orr M, Doellman D for Infusion Nurses Society. Infusion therapy standards of practice. *J Infus Nurs*. 2016; 39(Suppl 1):S1-S159

Gorski LA, Hadaway L, Hagle M et al for Infusion Nurses Society. Infusion therapy standards of practice. *Journal of Infusion Nursing*. 2021; 44(Suppl 1)

Guest JF, Keating T, Gould D, Wigglesworth N. Modelling the annual NHS costs and outcomes attributable to healthcare-associated infections in England. *BMJ Open*. 2020;10(1):e033367. <https://doi.org/10.1136/bmjopen-2019-033367>

Hadaway L. Short peripheral intravenous catheters and infections. *J Infus Nurs*. 2012;35(4):230-240. <https://doi.org/10.1097/NAN.0b013e31825af099>

Helm RE, Klausner JD, Klemperer JD, Flint LM, Huang E. Accepted but unacceptable: peripheral IV catheter failure. *J Infus Nurs*. 2015; 38(3): 189-203. <https://doi.org/10.1097/nan.0000000000000100>

Hibbard JS. Analyses comparing the antimicrobial activity and safety of current antiseptic agents: a review. *J Infus Nurs*. 2005;28(3):194-207. <https://doi.org/10.1097/00129804-200505000-00008>

Hovik LH, Gjeilo KH, Lydersen S, et al. Monitoring quality of care for peripheral intravenous catheters; feasibility and reliability of the peripheral intravenous catheters mini questionnaire (PIVC-miniQ). *BMC Health Serv Res*. 2019;19(1):636

Leong LEX, Lagana D, Carter GP, Wang Q, Smith K, Stinear TP, et al. *Burkholderia lata* infections from intrinsically contaminated chlorhexidine mouthwash, Australia, 2016. *Emerg Infect Dis*. 2018;24(11):2109-2111

Loveday HP, Wilson JA, Pratt RJ, et al. epic3: national evidence-based guidelines for preventing healthcare-associated infections in NHS hospitals in England. *J Hosp Infect*. 2014;86 Suppl 1:S1-S70

Maki DG, Kluger DM, Crnich CJ. The risk of bloodstream infection in adults with different intravascular devices: a systematic review of 200 published prospective studies. *Mayo Clin Proc*. 2006; 81:1159-1171

McDonald CP, Lowe P, Roy A et al. Evaluation of donor arm disinfection techniques. *Vox Sang*. 2001; 80(3):135-141. <https://doi.org/10.1046/j.1423-0410.2001.00029.x>

Medicines and Healthcare products Regulatory Agency. A guide to what is a medicinal product. Guidance Note 8. March 2020. <https://tinyurl.com/y8j7tu6t> (accessed 25 November 2020)

Mermel LA. Short-term peripheral venous catheter-related bloodstream infections: a systematic review. *Clin Infect Dis*. 2017;65(10):1757-1762. <https://doi.org/10.1093/cid/cix562>

Mestre G, Berbel C, Tortajada P et al. Successful multifaceted intervention aimed to reduce short peripheral catheter-related adverse events: a quasiexperimental cohort study. *Am J Infect Control*. 2013; 41(6):520-526. <https://doi.org/10.1016/j.ajic.2012.07.014>

Mimoz O, Lucet JC, Kerforne T et al. Skin antisepsis with chlorhexidine-alcohol versus povidone iodine-alcohol with and without skin scrubbing for prevention of intravascular-catheter-related infection (CLEAN): an open-label, multicentre, randomised, controlled, two by-two factorial trial. *Lancet*. 2015; 386(10008):2069-2077. [https://doi.org/10.1016/S0140-6736\(15\)00244-5](https://doi.org/10.1016/S0140-6736(15)00244-5)

Moureaux NL, Flynn J. Disinfection of needleless connector hubs: clinical evidence systematic review. *Nurs Res Pract*. 2015;2015:796762. <https://doi.org/10.1155/2015/796762>

Mutalib M, Evans V, Hughes A, Hill S. Aseptic non touch technique and catheter related blood stream infection in children receiving total parental nutrition at home. *United European Gastroenterology Journal*. 2015; 3(4), 393-398. <https://doi.org/10.1177/2050640615576444>

Nelson AM. Best practice in nursing: a concept analysis. *Int J Nurs Stud*. 2014;51(11):1507-1516. <https://doi.org/10.1016/j.ijnurstu.2014.05.003>

NHS Clinical Evaluation Team. Clinical review: safety peripheral intravenous cannula. 2018. <https://tinyurl.com/ybrjbcce> (accessed 25 November 2020)

Rickard CM, McCann D, Munnings J, McGrail MR. Routine resite of peripheral intravenous devices every 3 days did not reduce complications compared with clinically indicated resite: a randomised controlled trial. *BMC Med*. 2010;8:53. <https://doi.org/10.1186/1741-7015-8-53>

Rowley S, Clare S, Macqueen S, Molyneux R. ANTT v2: An updated practice framework for aseptic technique. *Br J Nurs*. 2010; 19(5 suppl): S5-S11. <https://doi.org/10.12968/bjon.2010.19.Sup1.47079>

Rowley S, Clare S. Right asepsis with ANTT® for infection prevention. In: Moureaux N (ed). *Vessel health and preservation: the right approach for vascular access*. Springer; 2019. <https://doi.org/10.1007/978-3-030-03149-7>

Rowley S, Clare S. How widely has ANTT been adopted in NHS hospitals and community care organizations in England and Scotland? *Br J Nurs*. 2020;29(16):924-932. <https://doi.org/10.12968/bjon.2020.29.16.924>

Safdar N, Maki DG. The pathogenesis of catheter-related bloodstream infection with noncuffed short-term central venous catheters. *Intensive Care Med*. 2004; 30(1):62-67. <https://doi.org/10.1007/s00134-003-2045-z>

Saliba P, Hornero A, Cuervo G et al. Interventions to decrease short-term peripheral venous catheter-related bloodstream infections: impact on incidence and mortality. *J Hosp Infect*. 2018; 100(3): e178-e186. <https://doi.org/10.1016/j.jhin.2018.06.010>

CPD reflective questions

- What are the potential harms and implications for patient safety when unlicensed medicinal products are used for skin antisepsis?
- How does the proprietary ANTT approach help improve the practice of aseptic technique generally, and skin antisepsis specifically?
- How does the design and application of skin antisepsis products affect the effectiveness of aseptic technique?

- Sato A, Nakamura I, Fujita H et al. Peripheral venous catheter-related bloodstream infection is associated with severe complications and potential death: a retrospective observational study. *BMC Infect Dis.* 2017;17(1):434. <https://doi.org/10.1186/s12879-017-2536-0>
- Silva P. The right skin preparation technique: a literature review. *J Perioper Pract.* 2014;24(12):283–285. <https://doi.org/10.1177/175045891402401204>
- Song JE, Kwak YG, Um TH et al. Outbreak of *Burkholderia cepacia* pseudobacteremia caused by intrinsically contaminated commercial 0.5% chlorhexidine solution in neonatal intensive care units. *J Hosp Infect.* 2018;98(3):295–299. <https://doi.org/10.1016/j.jhin.2017.09.012>
- Steere L, Ficara C, Davis M, Moureau M. Reaching one peripheral intravenous catheter (PIVC) per patient visit with lean multimodal strategy: the PIV5Rights™ bundle. *Journal of the Association for Vascular Access.* 2019;24(3):31–43. <https://doi.org/10.2309/j.java.2019.003.004>
- Tarka P, Chojecka A, Paduch O, Nitsch-Osuch A, Kanecki K, Kierzkowska A. Bactericidal activity of ready-to-use alcohol-based commercial wipes according to EN 16615 carrier standard. *Int J Environ Res Public Health.* 2019;16(18):3475. <https://doi.org/10.3390/ijerph16183475>
- Taylor JE, McDonald SJ, Earnest A et al. A quality improvement initiative to reduce central line infection in neonates using checklists. *Eur J Pediatr.* 2017;176(5): 639–646. <https://doi.org/10.1007/s00431-017-2888-x>
- Tepus D, Fleming E, Cox S, Hazelett S, Kropp D. Effectiveness of chloraprep in reduction of blood culture contamination rates in emergency department. *J Nurs Care Qual.* 2008;23(3):272–276. <https://doi.org/10.1097/01.ncq.0000324593.84213.4f>
- Trinh TT, Chan PA, Edwards O et al. Peripheral venous catheter-related staphylococcus aureus bacteremia. *Infect Control Hosp Epidemiol.* 2011;32(6):579–583. <https://doi.org/10.1086/660099>
- Tung A. Best practices for central line insertion. *Int Anesthesiol Clin.* 2013;51(1):62–78. <https://doi.org/10.1097/AIA.0b013e31827da437>
- Umscheid CA, Mitchell MD, Doshi JA, Agarwal R, Williams K, Brennan PJ. Estimating the proportion of healthcare-associated infections that are reasonably preventable and the related mortality and costs. *Infect Control Hosp Epidemiol.* 2011;32(2):101–114. <https://doi.org/10.1086/657912>
- Van Donk P, Rickard CM, McGrail MR, Doolan G. Routine replacement versus clinical monitoring of peripheral intravenous catheters in a regional hospital in the home program: A randomized controlled trial. *Infect Control Hosp Epidemiol.* 2009;30(9):915–917. <https://doi.org/10.1086/599776>
- Vigeant P, Loo VG, Bertrand C et al. An outbreak of *Serratia marcescens* infections related to contaminated chlorhexidine. *Infect Control Hosp Epidemiol.* 1998;19(10):791–794. <https://doi.org/10.1086/647728>
- Weber DJ, Rutala WA, Sickbert-Bennett EE. Outbreaks associated with contaminated antiseptics and disinfectants. *Antimicrob Agents Chemother.* 2007;51(12):4217–4224. <https://doi.org/10.1128/aac.00138-07>
- Webster J, Osborne S, Rickard CM, Marsh N. Clinically-indicated replacement versus routine replacement of peripheral venous catheters. *Cochrane Database Syst Rev.* 2019;1(1):CD007798. <https://doi.org/10.1002/14651858.cd007798.pub5>
- Wiemken TL. Skin antiseptics in healthcare facilities: is a targeted approach necessary? *BMC Public Health.* 2019;19:1158.
- Zhang L, Cao S, Marsh N, et al. Infection risks associated with peripheral vascular catheters. *J Infect Prev.* 2016;17(5):207–213
- Zingg W, Pittet D. Peripheral venous catheters: an under-evaluated problem. *Int J Antimicrob Agents.* 2009;34(4):S38–S42